Measuring particle fluxes and sinking rateshow can polyacrylamide gel sediment traps help? Ken Buesseler & Andrew McDonnell* Woods Hole Oceanographic Institution (& *ETH)





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Outline

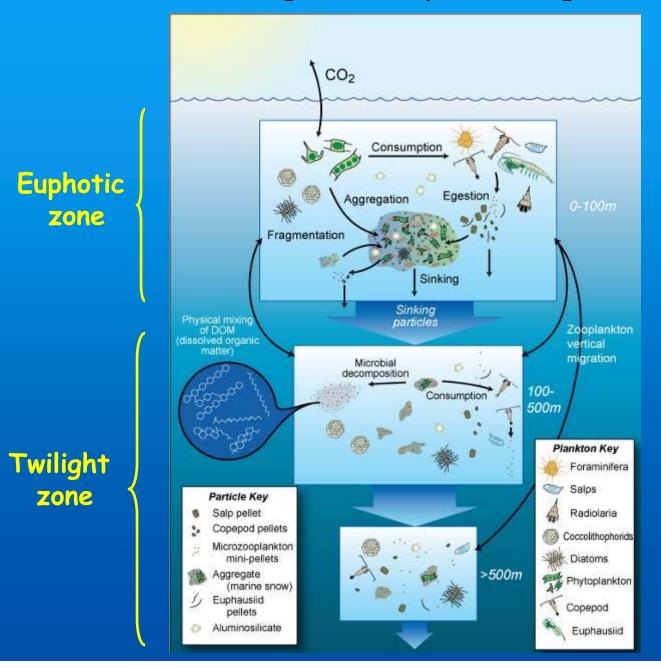
1. Biological pump- Motivation and current understanding 2. Particle sinking velocities- How do we determine?

3. Gel traps & water column particle studies

4. Future research directions



The Biological Pump & Twilight Zone



Combined processes which transfer organic matter and associated elements to depth

EZ = source of fresh sinking particles

TZ = layer of net loss of sinking POC

DEPTH MATTERS! *Buesseler & Boyd 2009*



Motivation behind biological pump & particle studies

Biological pump impacts surface to deep ocean DIC gradients and hence global C cycle and climate *Sarmiento and LeQuere*, 1996

Increase in remineralization depth by 25m will decrease atmos. CO₂ by ~20 ppm *Kwon et al. 2009*

>1 °C temp increase in twilight zone with climate change Levitus et al., 2009

Many elements (nutrients, TEI's) "hitch a ride on the bus" Gieskes, 1980 lecture SIO; Scavenging concept- Goldberg, 1954

The biological pump "feeds" the interior ocean and seafloor Alexander Agassiz, 1888

The twilight zone carbon budget is unbalanced Steinberg et al., 2008, Burd et al., 2010



Current understanding of biological pump & particle cycle

Global models do not adequately represent observed biogenic particle fluxes to the deep ocean Gehlen et al., 2006

No models have yet incorporated sufficient complexity to capture the observed variability of export fluxes *Boyd and Trull 2007*

The reason for this is we have not yet quantified the processes producing or transforming the particle flux Stemmann and Boss, 2011

The most critical parameter for particle flux is the particle settling speed in Stemmann and Boss, 2011 & attributed to Fasham et al., 1990

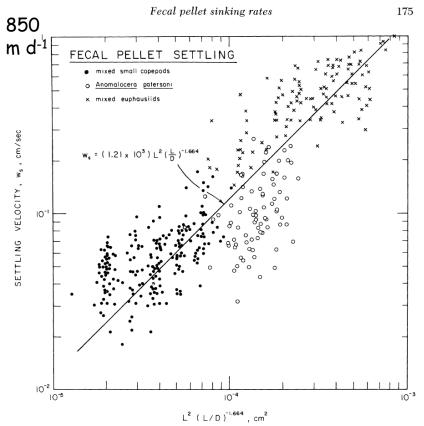


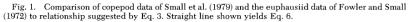
Given that it is so critical, how do we determine particle sinking speeds?

- 1. Settling columns
- 2. In-situ observations
- 3. Sediment trap peak matching
- 4. Settling velocity traps
- 5. Gel traps and particle imaging in water and flux



1. Settling columns





$$w_s = 0.0790 rac{1}{\mu} (
ho_s -
ho) g L^2 \Big(rac{L}{D}\Big)^{-1.664}$$

Limnol. Oceanogr., 26(1), 1981, 172-180

An analysis of sinking rates of natural copepod and euphausiid fecal pellets¹

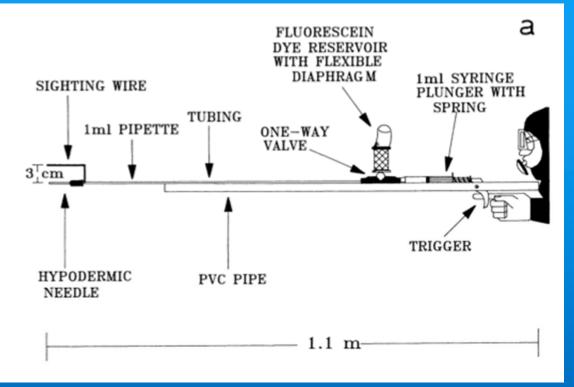
Paul D. Komar, Alan P. Morse, and Lawrence F. Small School of Oceanography, Oregon State University, Corvallis 97331

Scott W. Fowler International Laboratory of Marine Radioactivity,² Musée Océanographique, Prin

 lab vs. field vs. in situ?
 works best for large/fast intact particles pellets 100's-1000 m/d



2. In situ observations



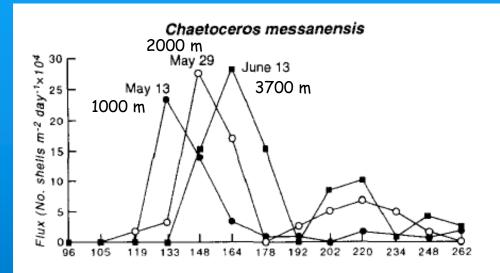
The human approach - limited to large, slow sinking marine snow - variable human limits

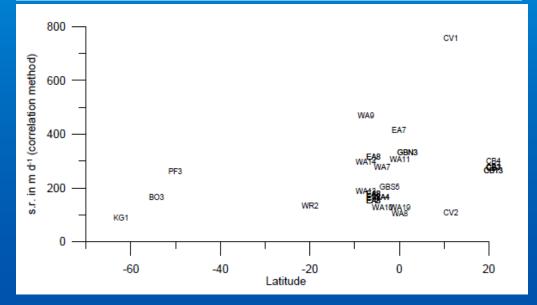
Alldredge and Gotschalk 1988

Progressed to in-situ work with cameras in trap tubes- Asper, Honjo et al. on ROVs- Silver, Pilskaln et al. on AUV's?



3. Sediment trap peak matching





NABE w = 200-250 m/d Honjo and Manganini, 1993

N. Atlantic- sinking rate & ballast, packaging relationships Fisher and Karakas, 2009

Sinking rate increases with depth see also Berelson 2002



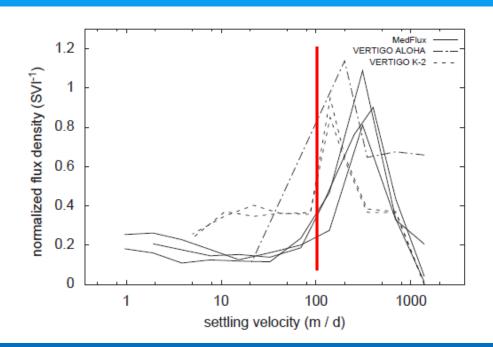




moored or drifting

LEE MI

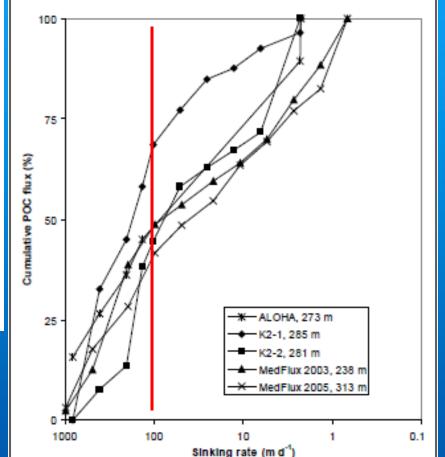
Sinking velocity trap- 2 examples 50% of flux > 100 m/d



MedFlux- Armstrong et al. 2009

Concerns

- it is still a trap
- changes in situ on rotating ball
- carry over between cups

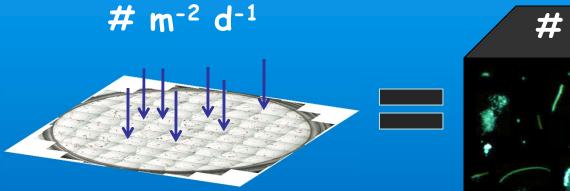


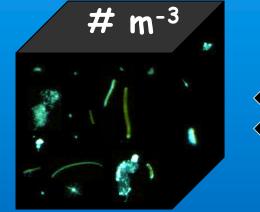
VERTIGO- Trull et al. 2008



5. Particle settling velocity from gel traps

$F_i = C_i \times W_{i,avg}$ Flux = Concentration × Avg. Sinking Velocity





m d-1



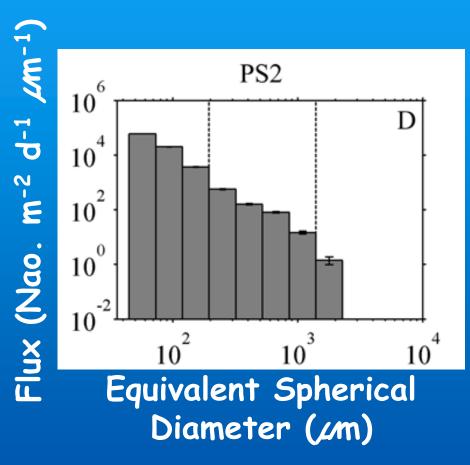


first used in McDonnell and Buesseler 2010



Measuring the flux size distribution

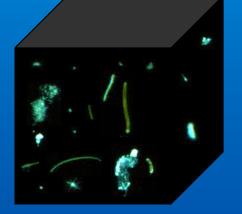






Measuring the concentration size distribution



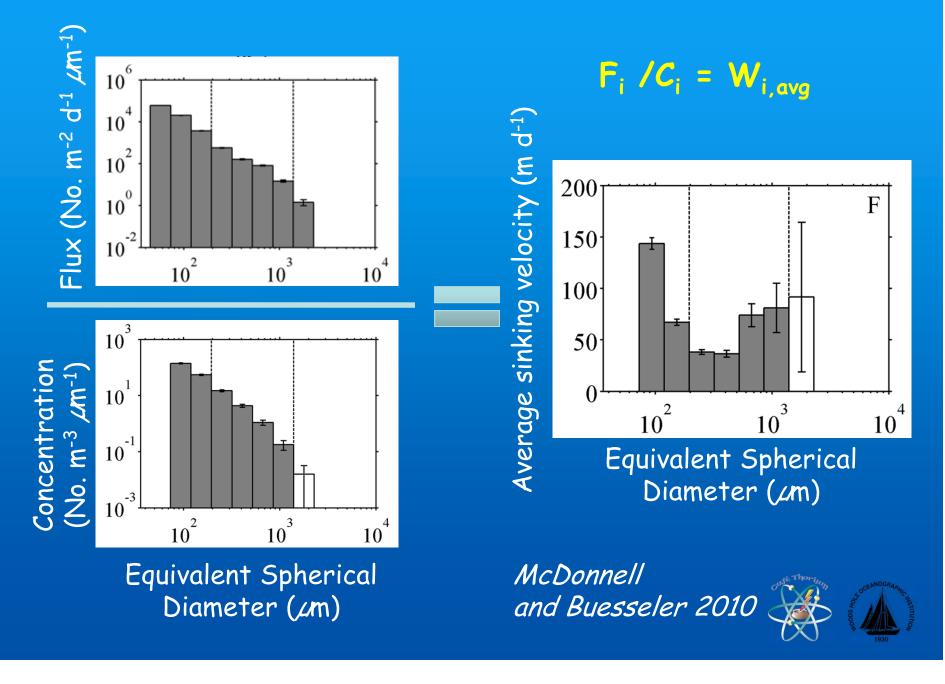


$10^{3} \underbrace{\qquad E \\ 10^{1} \\ 10^{-1} \\ 10^{-3} \underbrace{\qquad 10^{2} \\ 10^{2} \\ 10^{2} \\ 10^{3} \\ 10^{3} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\ 10^{4} \\$

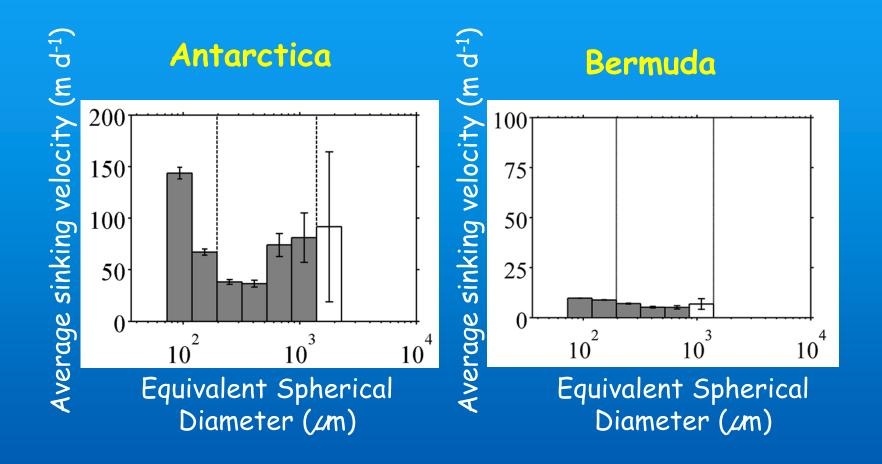
Equivalent Spherical Diameter (2m)



Calculating the average sinking velocity size distribution



Variability in regional average sinking velocities



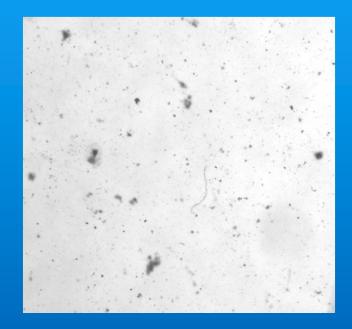


Sinking speed variability linked to differences in the particulate material

Antarctica

Bermuda



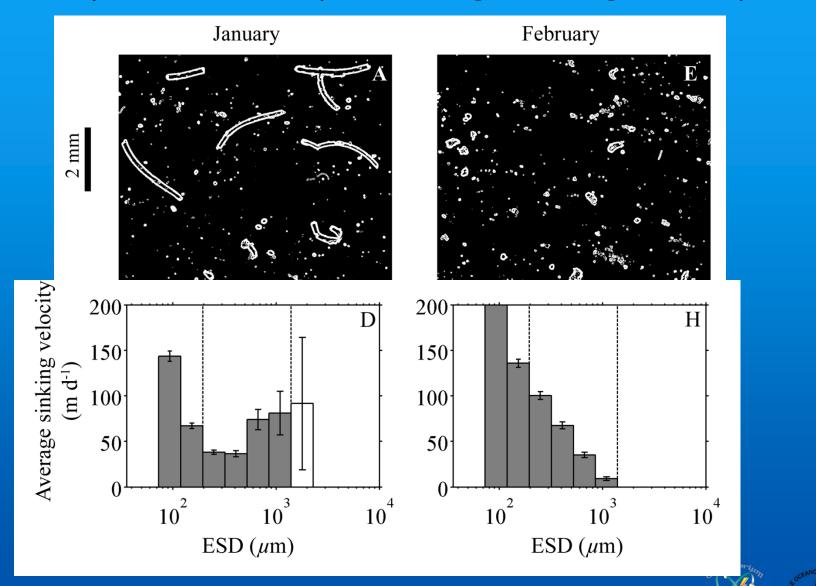


2 mm

* Gel traps quite useful for particle ID work see Waite and Nodder, 2001 Ebserbach et al, 2011

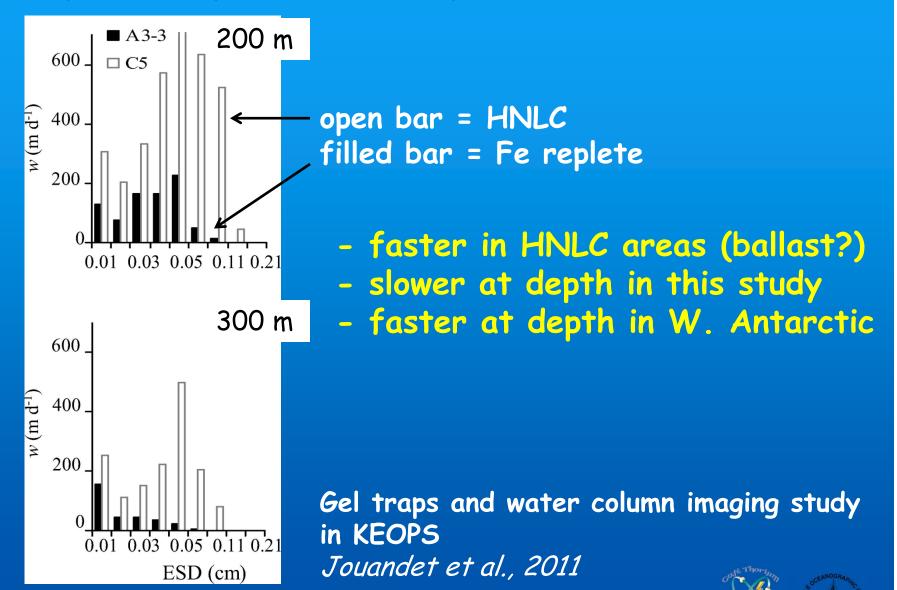


Temporal variability in average sinking velocity

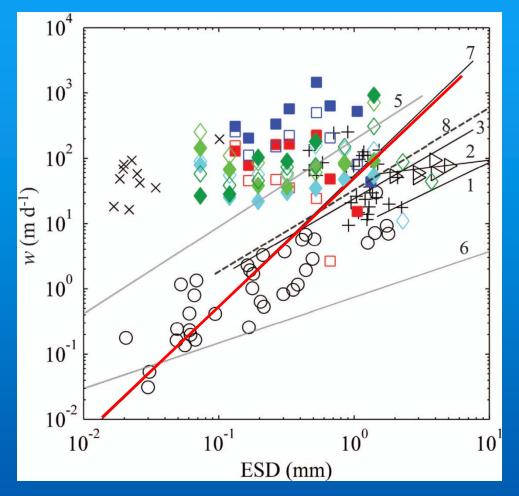


McDonnell and Buesseler 2010

Depth and spatial variability in sinking velocities



Can we put it all together?



Not very well....

Gel traps (color) other methods (B&W) #7 Stokes ≠ data No single relationship

Are we even measuring the same parameter? W_{individual} VS W_{average} What are limitations &

biases of each method?

Expect variability!

Jouandet et al., 2011 from Stemmann et al. 2004, Guidi et al. 2008 & many more



Grand challenges & future research directions

Need observations!

 multiple methods for flux, particle conc., sources, decomposition and sinking rates
 gel traps images provide source info
 gel traps and particle images provide sinking rates

Recognize variability exists on all space & time scales – moored inst., profilers, gliders can help resolve

BUT also include BIO in biopump PROCESS studies

- need to separate roles of zooplankton & bacteria

- physical controls on aggregation & biota linked

need to know processes to understand variability
 & predict changes in biopump due to climate

ASSUMING most critical parameter is sinking speed why does it vary?



